

体质量对雌雄孔雀鱼静止代谢率的影响*

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摘要:在 25 °C 下测定了 60 尾雌雄各半、不同大小的孔雀鱼(*Poecilia reticulata*)的静止代谢率(Resting metabolic rate)。结果表明:雄性孔雀鱼(体质量范围为 0.16~1.15 g)静止代谢率范围为 0.03~0.37 mg·h⁻¹;雌性孔雀鱼(体质量范围为 0.16~2.13 g)静止代谢率的范围为 0.03~0.44 mg·h⁻¹。协方差分析表明,孔雀鱼体质量对静止代谢率有显著影响($p < 0.05$),性别对静止代谢率的影响不显著,性别和体质量对静止代谢率的影响无交互作用。孔雀鱼的个体静止代谢率(A)与体质量(M)的关系可表示为: $\ln A = 0.858 \ln M - 0.643$ ($r^2 = 0.726, n = 60, p < 0.05$)。结果提示,尽管孔雀鱼发育非常迅速,但代谢尺度指数仍在文献报道范围内,体质量与代谢率的关系并无特殊性;孔雀鱼的特定体质量静止代谢率高于文献报道的其他鱼类,这可能与该鱼世代周期短、进化速度快有关。

关键词:孔雀鱼;体质量;代谢率

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代谢耗能是鱼类能量收支的一个重要组分,通常可划分为标准代谢(Standard metabolism)、运动代谢(Activity metabolism)、摄食代谢(Feeding metabolism)等组分。标准代谢是指外温动物在禁食、安静状态下的代谢,是维持基本生理功能的能量需求^[1-2]。由于难以完全保持鱼类身体处于绝对标准状态,在实验条件下测得的代谢率是标准代谢率的近似值,称为静止代谢率(Resting metabolic rate, RMR)^[3-4]。静止代谢率与鱼类的生理状态、社群地位、生活史对策等有关^[5-9]。

体质量是影响鱼类代谢率的重要因子。代谢率(R)随体质量(M)异速增加的关系可以表示为: $R = aM^b$,其中 a 为常数, b 为尺度指数, b 值的大小及变化规律反映了体质量影响代谢率的程度^[2]。代谢率随体质量增加的异速增长关系受到大量研究者的长期关注,已有广泛的研究和讨论^[10-15]。其中的一种重要理论基于分形几何学原理,对大量物种的代谢率数据进行分析,提出生物体的 b 值存在普适值 0.75^[16]。但研究表明,很多鱼类的 b 值并非 0.75^[17-23],有关鱼类的代谢率异速问题有待深入研究。

孔雀鱼(*Poecilia reticulata*),属鲈形目(Cyprinodontiformes)、花鳉科(Poeciliidae)、花鳉属(*Poecilia*),是一种繁殖周期短,性成熟快的鱼类,一般在出生后 50~60 d 即可达到性成熟。有关孔雀鱼的研究涉及生态学、进化、遗传学等领域^[24-26],而有关该鱼生物能量学方面的研究却鲜有报道。在孔雀鱼快速的发育过程中,代谢率随身体生长的变化可能具有典型的物种特异性。本研究对不同体质量的雌、雄孔雀鱼的静止代谢率进行了测定,初步探讨了体质量对该鱼静止代谢率的影响,旨在为生物能量学研究提供基础资料。

1 材料和方法

1.1 实验鱼的来源

孔雀鱼购自重庆市北碚区观赏鱼店,选择体表无明显伤害、无畸形的孔雀鱼于循环养殖箱中暂养,养殖水温为(25±1) °C,溶氧量接近饱和溶氧量^[2]。

1.2 实验方法

实验选取不同个体大小的雌、雄孔雀鱼各 30 尾,于流水式呼吸仪中测定代谢率^[27]。每个呼吸室放 1 尾鱼,

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另有 1 个呼吸室不放鱼作为空白,测定前调节呼吸室流速^[3]。驯化过夜后,于次日 9:00 开始测定耗氧率。采用溶氧仪(Microx TX3, PreSens-Precision Sensing GmbH Regensburg, Germany)测定溶氧量,并采用 5 mL 容量瓶收集水,以秒表记录历时,计算求得流速^[3]。静止代谢率(A ,单位: $\text{mg} \cdot \text{h}^{-1}$)计算方式为: $A = \Delta C_{\text{O}_2} \times v$; ΔC_{O_2} 为空白和实验鱼呼吸室溶氧量的差值($\text{mg} \cdot \text{h}^{-1}$), v 为呼吸室中水的流速($\text{L} \cdot \text{h}^{-1}$)。

1.3 数据处理方法

数据用“平均值±标准误”表示。采用 Excel 2003 对体质量与代谢率对数化后进行线性回归分析,用 SPSS Statistics 22.0 软件对体质量、性别及两者交互作用对代谢率的影响进行协方差分析,统计显著性水平为 $p < 0.05$ 。

2 结果

本研究中,雌雄孔雀鱼各 30 尾,在 25 °C 下,雄性孔雀鱼的体质量为(0.405 ± 0.047) g,范围为 0.16~1.15 g;个体静止代谢率为(0.11 ± 0.01) $\text{mg} \cdot \text{h}^{-1}$,范围为 0.03~0.37 $\text{mg} \cdot \text{h}^{-1}$;特定体质量代谢率为(289.1 ± 16.7) $\text{mg} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$,范围为 114.8~425.8 $\text{mg} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$ 。雌性孔雀鱼的体质量为(0.475 ± 0.092) g,范围为 0.16~2.13 g;个体静止代谢率为(0.12 ± 0.02) $\text{mg} \cdot \text{h}^{-1}$,范围为 0.03~0.44 $\text{mg} \cdot \text{h}^{-1}$;特定体质量代谢率为(264.5 ± 14.5) $\text{mg} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$,范围为 135.8~407.8 $\text{mg} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$ 。

采用对数直线回归对 25 °C 下雌雄孔雀鱼的个体静止代谢率(A)与体质量(M)的关系进行拟合,雄性、雌性孔雀鱼的有关拟合方程如图 1 中所示。协方差分析表明,体质量对个体静止代谢率有显著影响($p < 0.05$),但性别对个体静止代谢率的影响不显著,性别和体质量对个体静止代谢率也无交互作用,因此将雌鱼和雄鱼作为同一抽样进行回归分析,得到图 1 中所示的拟合方程。

3 讨论

随体质量增加,鱼类的个体代谢率增高^[4,28-30]。本实验中,雌雄孔雀鱼的个体代谢率均随体质量的增大而增高,二者间呈异速幂函数关系,与已有的研究结果一致。Jobling 总结多数鱼类的 b 值在 0.67~1.00 之间^[31],雌雄孔雀鱼的 b 值分别为 0.910 和 0.782,共同的 b 值为 0.858,均在文献报道的 b 值范围内。结果表明,尽管孔雀鱼发育非常迅速,但它的代谢率与体质量的关系并无明显的特殊性。

鱼的代谢水平会因性腺发育状况而变化^[32],代谢率的高低与性腺发育阶段有关^[33]。本研究中,孔雀鱼的性别对代谢率无明显影响,可能归因于雌性个体性腺发育程度不高。雌性孔雀鱼成熟的标志是臀鳍上前方的腹部出现黑色胎斑^[34],腹部膨大^[35]。本研究中的孔雀鱼尽管已经性成熟,但雌鱼腹部却没有膨大,反映了雌鱼较低的性腺发育程度。

采用文献报道的代谢率模型,分别计算出 25 °C 下 1 g 南方鲃(*Silurus meridionalis*)、鲃(*Silurus asotus*)、大鳍鱮(*Mystus macropterus*)和圆口铜鱼(*Coreius guichenoti*)特定体质量静止代谢率分别为 50.1^[4],63.9^[36],96.7^[37]和 96.5 $\text{mg} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$ ^[30]。在上述鱼类中,圆口铜鱼高于文献报道的多数物种^[30]。本研究中,根据方程求得 1 g 孔雀鱼的特定体质量静止代谢率为 227.3 $\text{mg} \cdot \text{kg}^{-1} \cdot \text{h}^{-1}$,远高于上述所有鱼类。孔雀鱼世代周期短,寿命只有 1~2 年^[35],进化速度极快^[38-39],可能与它的高代谢率有关,相关机制值得进一步研究讨论。

参考文献:

[1] 谢小军,孙儒泳.影响鱼类代谢的主要生态因素的研究进

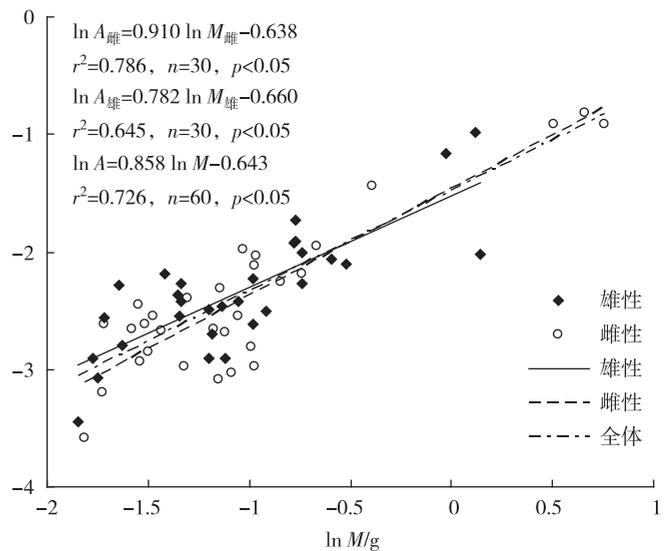


图 1 雌雄孔雀鱼体质量与静止代谢率的关系
Fig. 1 Relationships between body masses and resting metabolic rates of *P. reticulata*

展[J].西南师范大学学报:自然科学版,1989(4):141-149.

- Xie X J, Sun R Y. New perspective of the studies on several important ecological factors influencing metabolism of fishes[J]. Journal of Southwest China Normal University: Natural Science Edition, 1989(4):141-149.
- [2] 王文. 体重对圆口铜鱼代谢能力的影响[D]. 重庆: 西南大学, 2013.
- Wang W. Effects of body mass on metabolic capacity of *Coreius guichenoti* [D]. Chongqing: Southwest University, 2013.
- [3] 李格, 谢航, 何定聪, 等. 温度对斑马鱼静止代谢率和氨氮排泄率的影响[J]. 重庆师范大学学报: 自然科学版, 2015, 32(4):44-48.
- Li G, Xie H, He D C, et al. The effect of temperature on the metabolism and ammonia excretion in the zebrafish, *Danio rerio* [J]. Journal of Chongqing Normal University: Natural Science, 2015, 32(4):44-48.
- [4] Xie X J, Sun R Y. The bioenergetics of the southern catfish (*Silurus meridionalis*): 1. resting metabolic rate as a function of body weight and temperature[J]. Physiology Zoology, 1990, 63(6):1181-1195.
- [5] McNab B K. Food habits, energetics, and population biology of mammals[J]. The American Naturalist, 1980, 116(1):106-124.
- [6] McNab B K. The influence of food habits on the energetics of eutherian mammals[M]. Ecological Monographs, 1986, 56(1):1-19.
- [7] Metcalfe N B, Wright R J, Thorpe J E. Relationships between social status, otolith size at first feeding and subsequent growth in Atlantic salmon (*Salmo salar*) [J]. Journal of Animal Ecology, 1992, 61(3):585-589.
- [8] Metcalfe N B, Taylor A C, Thorpe J E. Metabolic rate, social status and life-history strategies in Atlantic salmon[J]. Animal Behaviour, 2010, 49(2):431-436.
- [9] Cutts C J, Metcalfe N B, Taylor A C. Aggression and growth depression in juvenile Atlantic salmon: the consequences of individual variation in standard metabolic rate [J]. Journal of Fish Biology, 2005, 52(5):1026-1037.
- [10] Oikawa S and Itazawa Y. Relative growth of organs and parts of the carp, *Cyprinus carpio*, with special reference to the metabolism-size relationship[J]. Copeia, 1984(3):800-803.
- [11] Kozłowski J, Konarzewski M, Gawelczyk A T. Cell size as a link between noncoding DNA and metabolic rate scaling [J]. Proceedings of the National Academy of Sciences of the United States of America, 2003, 100(24):14080-14085.
- [12] Agutter P S, Wheatley D N. Metabolic scaling: consensus or controversy? [J]. Theoretical Biology and Medical Modelling, 2004, 1(1):1-11.
- [13] Glazier D S. A unifying explanation for diverse metabolic scaling in animals and plants [J]. Biological Reviews, 2010, 85(1):111-138.
- [14] Kooijman S A L M. Dynamic energy budget theory for metabolic organisation[M]. Cambridge: Cambridge University Press, 2010.
- [15] Agutter P S, Tuszynski J A. Analytic theories of allometric scaling[J]. The Journal of Experimental Biology, 2011, 214(7):1055-1062.
- [16] West C B, Brown J H, Enquist B J. A general model for the origin of allometric scaling laws in biology[J]. Science, 1997, 276(5309):122-126.
- [17] Bokma F. Evidence against universal metabolic allometry [J]. Functional Ecology, 2004, 18(2):184-187.
- [18] Killen S S, Costa I, Brown J A, et al. Little left in the tank: metabolic scaling in marine teleosts and its implications for aerobic scope[J]. Proceedings of the Royal Society, 2007, 274(1608):431-438.
- [19] White C R, Cassey P, Blackburn T M. Allometric exponents do not support a universal metabolic allometry[J]. Ecology, 2007, 88(2):315-323.
- [20] Huang Q D, Zhang Y R, Liu S T, et al. Intraspecific scaling of the resting and maximum metabolic rates of the crucian carp (*Carassius auratus*) [J]. Plos One, 2013, 8(12):e82837.
- [21] Zhang Y R, Huang Q D, Liu S T, et al. Intraspecific mass scaling of metabolic rates in grass carp (*Ctenopharyngodon idellus*) [J]. Journal of Comparative Physiology B-Biochemical Systemic and Environmental Physiology, 2014, 184(3):347-354.
- [22] Luo Y P, He D C, Li G, et al. Intraspecific metabolic scaling exponent depends on red blood cell size in fishes[J]. The Journal of Experimental Biology, 2015, 218(10):1496-1503.
- [23] Li G, Xie H, He D C, et al. Effects of body chemical components on the allometric scaling of the resting metabolic rate in four species of cyprinids[J]. Fish Physiology and Biochemistry, 2016, 42(1):295-301.
- [24] Aich A, Chattopadhyay B, Datta, S, et al. Impact of composite tannery effluent on the amino-transferase activities in a fish biosystem, using guppy fish (*Poecilia reticulata*) as an experimental model[J]. Toxicological and Environmental Chemistry, 2011, 93(1):85-91.
- [25] 房英春, 齐跃, 李莹, 等. 盐酸环丙沙星、恩诺沙星和诺氟沙星对孔雀鱼急性毒性实验研究[J]. 沈阳大学学报: 自然科学版, 2012, 24(3):15-17.
- Fang Y C, Qi Y, Li Y, et al. Acute toxicity experience to guppy with ciprofloxacin HCl, enrofloxacin and norfloxacin[J]. Journal of Shenyang University: Nature Science, 2012, 24(3):15-17.
- [26] Bassar R D, Ferrler R, Lopezsepulcre A, et al. Direct and indirect ecosystem effects of evolutionary adaptation in the

- trinidadian guppy (*Poecilia reticulata*) [J]. The American Naturalist, 2012, 180(2): 167-185.
- [27] Luo Y P, Xie X J. Specific dynamic action in two body size groups of the southern catfish (*Silurus meridionalis*) fed diets differing in carbohydrate and lipid contents [J]. Fish Physiology and Biochemistry, 2009, 34(4): 465-471.
- [28] Zeuthen E. Oxygen uptake as related to body size in organisms [J]. The Quarterly Review of Biology, 1953, 28: 1-12.
- [29] 曹振东, 彭姜岚, 付世建. 体重对大鳍鲃力竭性运动后过量耗氧的影响 [J]. 重庆师范大学学报: 自然科学版, 2009, 26(4): 17-19.
- Cao Z D, Peng J L, Fu S J. Effect of body weight on excess post-exercise oxygen consumption in *Mystus macropterus* [J]. Journal of Chongqing Normal University: Natural Science, 2009, 26(4): 17-19.
- [30] Luo Y P, Wang Q Q. Effects of body mass and temperature on routine metabolic rate of juvenile largemouth bronze gudgeon *Coreius guichenoti* [J]. Journal of Fish Biology, 2012, 80(4): 842-851.
- [31] Tytler P, Calow P. Fish energetics: new perspectives [M]. Baltimore, Johns Hopkins University, 1985: 213-230.
- [32] Beamish F W H, Trippel E A. Heat increment: a static or dynamic dimension bioenergetic models? [J]. Transactions of the American Fisheries Society, 1990, 119(4): 649-661.
- [33] Evans D O. Temperature independence of the annual cycle of standard metabolism in the pumpkinseed [J]. Transactions of the American Fisheries Society, 1984, 113(4): 494-512.
- [34] 潘璠, 周凤建, 唐兰萍, 等. “观赏第一鱼”—孔雀鱼的养殖 [J]. 北京水产, 2006(5): 58-59.
- Pang P, Zhou F J, Tang L P, et al. “First ornamental fish”—guppy farming [J]. Journal of Beijing Fisheries, 2006(5): 58-59.
- [35] 张宇. 孔雀鱼的饲养及繁殖 [J]. 特种经济动植物, 2008, 11(7): 25-27.
- Zhang Y. Feeding and breeding guppies [J]. Special Economic Animal and Plant, 2008, 11(7): 25-27.
- [36] 杨振才, 谢小军, 孙儒泳. 鲃鱼的静止代谢及其与体重、温度和性别的关系 [J]. 水生生物学报, 1995, 19(4): 368-372.
- Yang Z C, Xie X J, Sun R Y. The resting metabolic rate of the common catfish (*Silurus asotus*) as a function of body weight, temperature and sex [J]. Acta Hydrobiologica Sinica, 1995, 19(4): 368-372.
- [37] 陈娟, 谢小军. 体质量及温度对大鳍鲃代谢的影响 [J]. 西南大学学报: 自然科学版, 2006, 31(4): 138-142.
- Chen J, Xie X J. Effect of body size and temperature on the metabolism of bagrid catfish, *Mystus macropterus* [J]. Journal of Southwest University: Natural Science, 2006, 31(4): 138-142.
- [38] Reznick D N, Shaw F H, Rodd F H, et al. Evaluation of the rate of evolution in natural populations of guppies (*Poecilia reticulata*) [J]. Science, 1997, 275(530): 1934-1937.
- [39] Ghalambor C K, Hoke K L, Ruell E W, et al. Non-adaptive plasticity potentiates rapid adaptive evolution of gene expression in nature [J]. Nature, 2015, 525(7569): 372.

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Effect of Body Mass on the Resting Metabolic Rate of Male and Female Guppy, *Poecilia reticulata*

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Abstract: The resting metabolic rate (RMR) of 60 guppies (*Poecilia reticulata*) individuals with different body mass were measured at 25 °C. The results showed that the RMR of males (body size rang 0.16 to 1.15 g) and females (body size rang 0.16 to 2.13 g) were 0.03 to 0.37 mg · h⁻¹ and 0.03 to 0.44 mg · h⁻¹, respectively. The RMR was significantly influenced by body mass ($p < 0.05$), rather than by neither gender nor the interaction effect between body mass and gender. The equation of the relationship between the individual RMR (A) and body mass (M) was $\ln A = 0.858 \ln M - 0.643$ ($r^2 = 0.726$, $n = 60$, $p < 0.05$). The results suggest that although guppy grows very rapidly, its scaling exponent is within the range reported by the previous literatures, without special relationship between M and A . The guppy has higher mass-specific metabolic rates than those of other fish reported in the literatures, which may be related with its short generation cycle and rapid evolution.

Key words: guppy; body mass; metabolic rate